



## Diversity of insects in Ciliwung Riverbank condet region (Rindam Jaya), South Jakarta, Indonesia

Yorianta Hidayat Sasaerila<sup>1</sup>, Yusuf Baskoro<sup>2</sup>, Dewi Elfidasari<sup>1\*</sup>

<sup>1</sup> Department of Natural Resource Management, Faculty of Science and Technology, University of Al Azhar Indonesia. Jl. Sisingamangaraja Kebayoran Baru, Selong, South Jakarta, Indonesia

<sup>2</sup> Department of Biology, Faculty of Science and Technology, University of Al Azhar Indonesia. Jl. Sisingamangaraja Kebayoran Baru, Selong, South Jakarta, Indonesia

### Abstract

Insects are animal with the highest diversity in the world and is one of the fauna that can live in various habitats and plays as important role in maintaining the stability of ecosystems both on land and water. The great role of insects for environment leads to research by using insects as polluted environment bioindicator. One of the polluted environment in Jakarta is Ciliwung River. Ciliwung River is one of polluted river caused by many society and industrial activity around the river. Previous research of insects has been done in Srengseng Sawah and Manggarai Water Gate when Ciliwung River are not yet restored into sustainable condition. A preliminary research has been conducted in the Ciliwung River, Condet Region (Rindam Jaya), South Jakarta which aims to determine the diversity of insects in the area. This study conducted as preliminary data for the further insect's research of the Ciliwung River. The result showed that 390 individual insects were found with Formicidae family most widely found (227 individuals). This study also measured diversity values using frequency of attendance. The highest frequency of attendance score is in groups of Formicidae family insects (58.2%).

**Keywords:** Insects, Ciliwung River, pollution, diversity

### Introduction

Indonesia is a country with high biodiversity, as it has ecosystems, species within ecosystems and genetic traits in each species are very diverse. Therefore, Indonesia is known as mega biodiversity <sup>[1, 2]</sup>. This is because Indonesia is located in the tropics with a stable climate, and is an archipelagic country geographically located between the continents of Asia and Australia <sup>[3]</sup>.

Insects are one of the animal group with the highest diversity in the world with a 58% percentage in biodiversity globally. Insects can live in a variety of habitats and play an important role in maintaining the stability of ecosystems, both on land and water. Insects have modified and adaptable bodies capable of producing a variety of diversity capable of living in land and water environments. The diversity includes: the state and stability of the basic pattern is long-lasting also the variety is high (there are 29 orders found today). The number of insects individual reaches one quintillion (10<sup>18</sup>). <sup>[4, 5]</sup> The current number of insects species found reached 1.004.898 species. <sup>[6, 7, 8]</sup> Insects are known to have evolved beyond the evolution of land plants in the Silurian and Devonian periods of more than 400 million years ago. Collembola and apterygota insects show evidence of their relics in the Devonian period, while the flying insects in the Carbon period. <sup>[9, 10, 11]</sup>

Attention to environmental issues has increased the demand for bioindicators that can determine the conditions of the environment. <sup>[12, 13]</sup> Insects are the organism that can contribute as a general appraiser of an environmental sustainability level. Insect indicators are especially useful because more than half of all species and their diversity are able to assess differences in habitat on an acceptable scale.

<sup>[14, 15, 16]</sup> One of the insects used as bioindicators is Formicidae family (ants).

Research about the diversity of insects in the Ciliwung River was first conducted by Ruslan (2008) on the diversity of ground insects in the Ciliwung River region at two locations (Srengseng Sawah and Manggarai Water Gate) in Jakarta. <sup>[17]</sup> Ciliwung River is one of the rivers that play an important role in the life of the people of Jakarta. This river comes from Mount Pangrango, which flows through the Bogor Regency, Bogor City, Depok City and then to Jakarta. The condition of the Ciliwung River is now polluted due to the large number of community activities and industry around the river. <sup>[18, 19, 20]</sup> This causes a decline in the quality of Ciliwung River water, both used for human daily needs, and wildlife life in the habitat. This polluted condition is thought to affect the diversity of fauna, one of which is the diversity of insects. <sup>[21, 22]</sup>

This study was conducted prior to the massive cleanup of Ciliwung River which was led by Jakarta Military Command (Kodam Jaya). Further research is needed on the diversity of insects after Ciliwung River is cleaned.

### Material and Methods

#### 1. Research sites

This study was conducted on May – July 2017. The study consisted of sampling at 30 points along the Ciliwung River in Condet (Rindam Jaya) South Jakarta area and the sample analysis was conducted at the Biology Laboratory of Al Azhar University Indonesia and Cibinong Indonesian Academy of Science (LIPI Cibinong). Research procedure includes location survey, insect collection, sample identification, sample confirmation and diversity calculation using frequency of attendance.

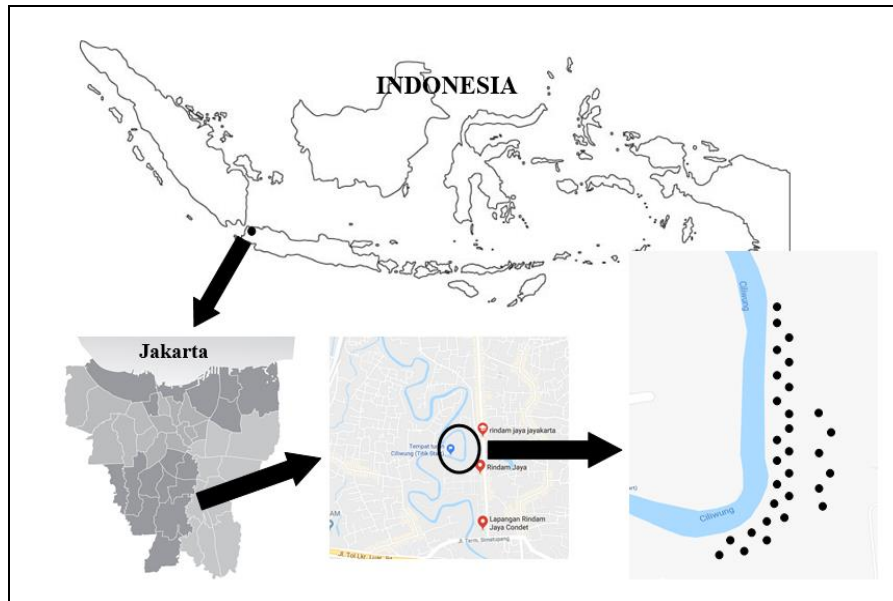


Fig 1: The insects sampling location with 30 dots of trap placement

**2. Material**

The material used three types of traps, Pitfall Trap, Yellow Pan Trap (YPT) and insect net, samples inside plastic seals, the sample bottle

**3. Procedures**

**3.1 Location survey**

Location survey are conducted from Rindam Jaya, Condet, South Jakarta to Bidara Cina using LCR (Landing Craft Rubber) boat along the Ciliwung River to determine the location where the traps are installed. Location were obtained at Rindam Jaya, Condet, South Jakarta.

**3.2 Insect collecting**

The installation of insect trap is done after determining the location of trap placement. We used three types of traps, Pitfall Trap to capture the ground insect, Yellow Pan Trap (YPT) to capture small flying insect and insect net to catch large flying insect.

**3.3 Sample identification**

All samples were taken to Biology Laboratory of Al Azhar University Indonesia for identification. Before samples were identified, samples inside plastic seals was transferred first to the sample bottle. Subsequently all insects were identified to the level of the Order. The identified samples are then recorded by number, type of Order, sample location, sampling time and sample size.

**3.4 Sample confirmation**

The identified samples were then taken to Indonesian Institut of Science or Lembaga Ilmu Pengetahuan Indonesia (LIPI) Cibinong for confirmation until Order level that had previously been done and further identification up to the Family level.

**4. Data analysis**

The samples that have been identified then calculated using Frequency of Attendance with the following formula:

$$FK = \frac{a}{n} \times 100\%$$

Note:

a= the number of collected family

n= total samples

- Description of results:
- 0-25% = very rare
  - 25-50% = uncommon
  - 50-75% = common
  - >75% = very common

**Result and Discussion**

The total number of individuals collected in this study were 390 specimens consisting of 7 orders, 1 Lepidoptera sample, 2 Blattodea samples, 19 Orthoptera samples, 16 Hemiptera samples, 55 Coleoptera samples, 235 Hymenoptera samples and 53 Diptera samples (Table 1).

Table 1: Data of insect samples collected (order and family) and Frequency of Attendance of each family

No.	Order	Family	Number of individuals	Frequency of attendance (FoA) (%)
1	Lepidoptera	Hesperiidae	1	0.26
2	Blattodea	Blattidae	2	0.51
3	Orthoptera	Acrididae	7	1.79
		Gryllidae	12	3.07
4	Hemiptera	Membracidae	16	4.2
5	Coleoptera	Carabidae	4	1.02
		Staphylinidae	51	13.1
6	Hymenoptera	Formicidae	227	58.2

		Crabronidae	1	0.26
		Figitidae	1	0.26
		Sphecidae	1	0.26
		Collitidae	1	0.26
		Icheumonidae	2	0.51
		Apidae	2	0.51
7	Diptera	Dolicophodidae	43	11.02
		Ephyridae	5	1.28
		Calliphoridae	1	0.26
		Tachinidae	4	1.02
8	Odonata	Libellulidae	7	1.79
		Coenagrionidae	2	0.51
Total individual			390	100

Previous study on two locations in Ciliwung River (Srengseng Sawah and Manggarai) showed that in Srengseng Sawah captured 561 Hymenoptera samples, 21 Orthoptera samples, 68 Coleoptera samples, and 15 Diptera samples. In Manggarai insect captured were 168 Hymenoptera samples, 5 Orthoptera samples, 37 Coleoptera samples and 20 Diptera samples. [17] The most commonly found family is the ant (Formicidae family). This is due to the high survival of ants. Ants can be

found all over the world except Iceland, Greenland, Antarctica, parts of Polynesia, and some remote islands in the Atlantic Ocean and India. Ants can also live in almost all habitats and terrestrial environments including deserts, beaches, inside walls and unused waterways. Some species are even able to live in the water for 14 days by turning into a sleeping state (dormant) that causes the reduction of oxygen required by ants twentyfold. [23, 24] The number of individuals found was 227 (FoA 58.2%).

**Table 2:** Collected insect data comparison between Rindam Jaya and study by Ruslan (2008) in Srengseng Sawah and Manggarai.

No.	Order	Rindam Jaya		Srengseng Sawah		Manggarai	
		Σ F	Σ I	Σ F	Σ I	Σ F	Σ I
1	Lepidoptera	1	1	0	0	0	0
2	Blattodea	1	2	0	0	0	0
3	Orthoptera	2	19	3	21	2	5
4	Hemiptera	1	16	1	1	1	1
5	Coleoptera	2	55	7	68	4	37
6	Hymenoptera	7	235	4	561	4	168
7	Diptera	4	53	6	15	6	20
8	Odonata	2	9	0	0	0	0
9	Homoptera	0	0	1	2	0	0
10	Psocoptera	0	0	1	1	1	1
11	Isoptera	0	0	1	1	0	0
Total		20	390	24	670	18	232

**Note:** ΣF is total number of Family and ΣI is total number of Individuals.

Formicidae family (ants) are generally used for soil quality bioindicators and have an important role in the improvement of degraded areas and forests under reforestation. This group of insects is very sensitive to human influences, and can be used as an environmental indicator in different ecosystems. [25, 26, 27] Many species of ants may become extinct somewhere depending on how much the environment is changing. This may lead to superior dominant species that can be used as indicators in an impaired environment. [28, 29] Until now there has been no research conducted to see the benefits of ants on the Ciliwung River.

The highest number of family collected from Hymenoptera order is from Rindam Jaya, while the highest number of family collected from Coleoptera order is from Srengseng Sawah. In the Srengseng Sawah area, 561 individuals of the Hymenoptera Order were found higher than the other two locations, but the Lepidoptera, Blattodea and Odonata Order were not found in Srengseng Sawah and Manggarai (Table 2). This can be caused by several factors, one of which is the abiotic factor. One of the abiotic factors affecting dragonflies (Odonata order) is water pH although it does not affect significantly. [30, 31, 32]

Based on research conducted by Elfidasari *et al* (2020) the pH of Ciliwung River's water is 6.9. [33] The existence of dragonfly is quite tolerant where in water with a very acidic pH (<4.00) there are still certain types of dragonflies, although in very small quantities, which the dragonfly can not develop in neutral waters. However, waters with a pH below 3.00 can cause damage to dragonflies. [34, 35] It can be explained that dragonflies can be found in the Ciliwung River due to the dragonfly is quite tolerant even at very low pH, while the pH of Ciliwung River is neutral.

For the Hesperidae Family (Lepidoptera order) only one specimen was obtained because the method of catching using insect net was less efficient (only one to two people who capturing with insect net). Crabronidae family, Figitidae family, Sphecidae family, Colletidae family (Hymenoptera order) and Calliphoridae family (Diptera order) also get only one specimen from each family because the method used is not specific to the insects family. The Frequency of Presence (FoA) of this family is only 0.2% (Table 1). The method used is Yellow Pan Trap (YPT), which is a common method used to capture parasitoid insects. [36, 37] Based on the recommendation of an insect researcher at LIPI Cibinong, trap was filled with water and was given a drop of dish soap, and trap was placed for only

few hours (8 am – 12 pm). This short term trap placement was to prevent the insect for being damaged and may cause only a few insects to be caught.

Butterflies and moths (Lepidoptera order) live in temperate and tropical climates and very sensitive to environmental changes. [38, 39] Several groups of Lepidoptera are used as

indicators of environmental pollution of heavy metals and carbon dioxide at locations adjacent to industrial and residential areas. The presence of bronze, iron, nickel, cadmium, sulfuric acid ions and other substances used in fertilizers were studied with pupae of different species from Geometridae and Noctuidae family. [24]



**Fig 1:** Environmental condition of sampling location

The Coleoptera order represents about 20% of the total arthropod diversity and plays a role in maintaining soil quality, regulation of invertebrate populations, and contributes to soil physical and chemical content. [40] Beetles from the Coleoptera Order and the Carabidae Family are important predators participating in biological control and

biological monitoring of pollution from oil, sulfur, herbicide, CO<sub>2</sub>, insecticides and radioactive phosphorus. [41, 42] The beetle family (Coleoptera: Scarabidae) has a high potential as an environmental indicator in forest and agricultural areas. [43]

**Table 3:** Bioindicator insect groups in aquatic and terrestrial environments and their role in environmental biomonitoring. [44]

Group	General Name	Biomonitoring	Habitat
Odonata Order	Anisoptera & Zygoptera	Water quality	Aquatic
Grynidae Dysticidae Hydrophilidae Notonectidae Famili Vellidae	Whirligig beetles Predaceous diving beetles - Backswimmer -	Highly adaptable	Aquatic
Ephemeroptera Plecoptera Order	Mayflies Stoneflies	Highly adaptable	Aquatic
Halobates	Ocean-skaters	Cadmium and lead	Aquatic
Coleoptera order Scarabaeidae family	Beetle	Forest and agriculture	Land
Ordo Coleoptera Famili Carabidae	Beetle	Biological oil control, sulfur, herbicide, CO <sub>2</sub> , insecticide pollution	Land
Lepidoptera order	Moth and butterfly	More sensitive with environmental change with heavy metal and CO <sub>2</sub> pollution	Land
Collembola order	Springtails	Heavy metal pollution, pesticide and water acidity	Land
Formicidae family	Ants	Reforestation	Land
Diptera order Sarcophagidae family	Flies and mosquito	Heavy metal	Land
Diptera order Syrphidae family	Flies and mosquito	Affected by decreased diversity	Land
Apis melifera	Domestic bees	Chemical environmental change	Land

The Frequency of Attendance (FoA) of the Staphylinidae family (Coleoptera order) is quite large compared to other families (13%) (Table 1). This is presumably because Staphylinidae can live in various environments such as under rocks and other objects on the ground, and can live on fungi and leaf remains. [45] (Borror *et al.* 1992), in which the majority of sampling environment location is soil mixed with rocks and many other objects on the ground (Figure 1). Diptera order is a very heterogeneous insect group but is still limited in its use as a bioindicator due to the lack of ecological knowledge of many flies. The fly species from the Sarcophagidae family have potential as indicators of environmental pollution such as heavy metals, asbestos fibers and chemical wastes. [41, 46] It is recommended that the use of flies (Diptera order) as a chemical indicator of

contaminated soil should be careful because of the variability of flies sensitivity to insecticides and herbicides. [41, 47, 48] In this order, the Dolichopodidae family has a highest number on study location (11%) (Table 1). The Dolichopodidae family (Diptera order) is a family of flies that can live in many places, especially near swamps, waterways, forests and grasslands, but many of these flies can only live in certain environments. The larvae are present in mud or water, on decomposed wood, under bark and on grass stems. [45, 49] (Borror *et al.* 1992). Many of the samples obtained at the study location were bioindicator insects such as Odonata, Coleoptera, Lepidoptera and Diptera, as well as the Carabidae (Ordo Coleoptera) family and the Formicidae Family (Hymenoptera order). [21, 24, 41] This is appropriate as

presented in the research of da Rocha *et al* (2010) that the insect groups collected are included in environmental bioindicators (Table 3).<sup>[44]</sup>

### Conclusion

The result showed that 390 individual insects were found with Formicidae family most widely found (227 individuals). The highest frequency of attendance score is in groups of Formicidae family insects (58.2%).

### Acknowledgment

The author would like to thank the PT. Perusahaan Gas Negara (PGN) for funding the research activities, as well as TNI Jakarta Military District Command (Kodam Jaya) especially Lt. Gen. Agus Sutomo (Former Commander of Kodam Jaya), Maj. Gen. Teddy Lhaksamana (Former Commander of Kodam Jaya), Col. Inf. Arudji Anwar (Aster Kasdam Jaya), Lt. Col. Czi. Edi Martadinata, Lt. Col. Inf. Firdaus Agustiana (South Jakarta Dandim) and TNI personnel who accompanying author in the field and LCR operators (Serda Apwaris, Kophka Ahlan, Pelda Yus Setiawan, Setyohadi and Praka Mulyanto), also Rindam Jaya Provos who has given permission to conduct research in Rindam Jaya area.

### References

- Sanka I, Kusuma AB, Martha F, Hendrawan A, Pramanda IT, Wicaksono A, *et al*. Synthetic biology in Indonesia: Potential and projection in a country with mega biodiversity. *Biotechnology Notes*,2023;4:41-48. doi: <https://doi.org/10.1016/j.biotno.2023.02.002>
- Suyadi S, Nugroho DA, Irawan A, Pelasula D, Ruli F, Islami MM, *et al*. Biodiversity in the coastal ecosystems of small islands and its conservation status. *IOP Conference Series: Earth and Environmental Science*,2021;762012024. doi: 10.1088/1755-1315/762/1/012024
- Munawar M, Pasetya TAE, McNeil R, Jani R, Buya S. Spatio and Temporal Analysis of Indonesia Land Surface Temperature Variation During 2001-2020. *Journal of The Indian Society of Remote Sensing*,2023;51(7):1393-1407. doi: 10.1007/s12524-023-01713-0
- Raghavendra KV, Bhoopathi T, Gowthami R, Keerthi MC, Suroshe SS, Ramesh KB, *et al*. Insects: biodiversity, threat status and conservation approaches. *Current Science*,2022;122(12):1374-1384. doi: <https://doi.org/10.18520/cs/v122/i12/1374-1384>
- Wagner DL, Fox R, Salcido DM, Dyer LA. A window to the world of global insect declines: Moth biodiversity trends are complex and heterogeneous. *PNAS*,2021;118(2):e2002549117. doi: <https://doi.org/10.1073/pnas.2002549117>
- Cardoso P, Leather SR. Predicting a global insect apocalypse. *Insect Conservation and Diversity*,2020;12(4):263-267. doi: <https://doi.org/10.1111/icad.12367>
- Raven PH, Wagner DL. Agricultural intensification and climate change are rapidly decreasing insect biodiversity. *PNAS*,2021;118(2):e2002549117. doi: <https://doi.org/10.1073/pnas.2002549117>
- Forister ML, Pelton EM, Black SH. Declines in insect abundance and diversity: We know enough to act now. *Conservation Science and Practice*,2019;1(8):e80. doi: <https://doi.org/10.1111/csp.2.80>
- Cannell A, Blamey N, Brand U, Escapa I, Large R. A revised sedimentary pyrite proxy for atmospheric oxygen in the Paleozoic: Evaluation for the Silurian-Devonian-Carboniferous period and the relationship of the results to the observed biosphere record. *Earth-Science Reviews*,2022;231:104062. doi: <https://doi.org/10.1016/j.earscirev.2022.104062>
- Tihelka E, Howard RJ, Cai C, Lozano-Fernandez J. Was there a Cambrian explosion on land? The case of arthropod terrestrialization. *Journals Biology*,2022;11(10):1516. doi: <https://doi.org/10.3390/biology11101516>
- Prokop J, Nel A, Engel MS. Diversity, form, and postembryonic development of Paleozoic insects. *The Annual Review of Entomology*,2023;68:401-429. doi: <https://doi.org/10.1146/annurev-ento-120220-022637>
- Elizalde L, Arbetman M, Arnan X, Eggleton P, Leal IR, Lescano MN, *et al*. The ecosystem services provided by social insects: traits, management tools and knowledge gaps. *Biological Review*,2020;95(5):1418-1441. doi: <https://doi.org/10.1111/brv.12616>
- Eggleton P. The state of the world's insects. *Annual Review of Environment and Resources*,2020;45:61-82. doi: <https://doi.org/10.1146/annurev-enviro-012420-050035>
- Kovac M, Gasparinni P, Notarangelo M, Rizzo M, Canellas I, Fernandez-de-Una L, *et al*. Towards a set of national forest inventory indicators to be used for assessing the conservation status of the habitats directive forest habitat types. *Journal of Nature Conservation*,2020;53:125747. doi: <https://doi.org/10.1016/j.jnc.2019.125747>
- Zaghloul A, Saber M, Gadow S, Awad F. Biological indicators for pollution detection in terrestrial and aquatic ecosystems. *Bulletin of the National Research Centre*,2020;44:127. doi: <https://doi.org/10.1186/s42269-020-00385-x>
- Prendergast KS, Dixon KW, Bateman PW. A global review of determinants of native bee assemblages in urbanised landscapes. *Insect Conservation and Diversity*,2022;15(4):385-405. doi: <https://doi.org/10.1111/icad.12569>
- Ruslan H. A study on the diversity of soil surface insects in the riparian areas of the Ciliwung River at two locations (Srengseng Sawah and Pintu Air Manggarai) in Jakarta. *Jurnal Ilmu dan Budaya*,2008;28(11):1032-1040.
- Atmanto D. Model of river environmental management based on sustainable community social capital. *AIP Conference Proceeding*,2024;2982:040028. doi: <https://doi.org/10.1063/5.0185176>
- Mahendra APD, Pratama MA, Moersidik SS, Rahmawati S, Iresha FM. Spatial dynamics of microplastic pollution in water and sediments of the Ciliwung river along with conditions of water Quality field parameters and population density. *Journal of Ecological Engineering*,2023;24(8):296-309. doi: <https://doi.org/10.12911/22998993/166311>
- Elfidasari D, Ismi LN, Sugoro I. Heavy Metals Concentration in Water, Sediment, and Pterygoplychthys pardalis in the Ciliwung River, Indonesia. *AACL-Bioflux*,2020;13(3):1764-1778.
- Parikh G, Rawtani D, Khatri N. Insects as an indicator for environmental pollution. *Environmental Claims*

- Journal,2021:33(2):161-181. doi: <https://doi.org/10.1080/10406026.2020.1780698>
22. Chowdhury S, Dubey VK, Choudgury S, Das A, Jeengar D, Sujatha B, *et al.* Insects as bioindicator: A hidden gem for environmental monitoring. *Frontiers Environmental Science*,2023;11:1-16. doi: <https://doi.org/10.3389/fenvs.2023.1146052>
  23. Chen C, Mahar R, Merritt ME, Hahn DA. ROS and hypoxia signaling regulate periodic metabolic arousal during insect dormancy to coordinate glucose, amino acid, and lipid metabolism. *PNAS*,2021;118(1):e2017603118. doi: <https://doi.org/10.1073/pnas.2017603118>
  24. Heliovaara K. *Insects and Pollution*. 1st Edition,2017, Boca Raton. Pages 407. doi: <https://doi.org/10.1201/9781351073622>
  25. Segat JC, Vasconcellos RLF, Silva DP, Baretta D, Cardoso EJBN. Ants as indicators of soil quality in an on-going recovery of riparian forests. *Forest Ecology and Management*,2017;404:338-343. doi: <https://doi.org/10.1016/j.foreco.2017.07.038>
  26. Nsengimana V, Dekoninck W. Soil-litter ant (Hymenoptera: Formicidae) community response to reforested lands of Gishwati tropical montane forest, northern-western part of Rwanda. *Journal of Tropical Ecology*,2021;37(4):165-174. doi: <https://doi.org/10.1017/S0266467421000237>
  27. Ribeiro LG, Silva AO, Vas KA, dos Santos JV, Nunes CA, Carneiro MAC. Soil arthropod community responses to restoration in areas impacted by iron mining tailings deposition after Fundão dam failure. *Environmental Monitoring Assessment*,2023;195:1299. doi: <https://doi.org/10.1007/s10661-023-11843-0>
  28. Alroy J. Effects of habitat disturbance on tropical forest biodiversity. *PNAS*,2017;114(23):6056-6061. doi: <http://www.pnas.org/cgi/doi/10.1073/pnas.1611855114>
  29. Sullivan GT, Ozman-Sullivan SK. Alarming evidence of widespread mite extinctions in the shadows of plant, insect and vertebrate extinctions. *Austral Ecology*,2020;46(1):163-176. doi: <https://doi.org/10.1111/aec.12932>
  30. Buczynska E, Buczynski P. Survival under anthropogenic impact: the response of dragonflies (Odonata), beetles (Coleoptera) and caddisflies (Trichoptera) to environmental disturbances in a two-way industrial canal system (central Poland). *PeerJ*,2019;6:e6215. doi: <https://doi.org/10.7717/peerj.6215>
  31. Bastos RC, Brasil LS, Oliveira-Junior JMB, Carvalho FG, Lennox GD, Barlow J, Juen L. Morphological and phylogenetic factors structure the distribution of damselfly and dragonfly species (Odonata) along an environmental gradient in Amazonian streams. *Ecological Indicators*,2021;122:107257. doi: <https://doi.org/10.1016/j.ecolind.2020.107257>
  32. Oliveira-Junior JMB, Juen L. Structuring of Dragonfly Communities (Insecta: Odonata) in Eastern Amazon: Effects of Environmental and Spatial Factors in Preserved and Altered Streams. *Insects*,2019;10(10):322. doi: <https://doi.org/10.3390/insects10100322>
  33. Elfidasari D, Wijayanti F, Muthmainah HF. Short Communication: The effect of water quality on the population density of *Pterygoplichthys pardalis* in the Ciliwung River, Jakarta, Indonesia. *Biodiversitas*,2020;21(9):4100-4106. doi: <https://doi.org/10.13057/biodiv/d210922>
  34. Perron MA, Pick FR. Water quality effects on dragonfly and damselfly nymph communities: A comparison of urban and natural ponds. *Environmental Pollution*,2020;263:114472. doi: <https://doi.org/10.1016/j.envpol.2020.114472>
  35. Ilhamdi ML, Al Idrus A, SantodoD, Hadiprayitno G, Syazali M. Species richness and conservation priority of dragonflies in the Suranadi Ecotourism Area, Lombok, Indonesia. *Biodiversitas*,2021;22(4):1846-1852. doi: <https://doi.org/10.13057/biodiv/d220430>
  36. Bauer L, Hansen J, Gould J. *Yellow pan traps: A simple method for trapping parasitoids released for biological control of the Emerald Ash Borer*. Department of Entomology Michigan State University. Michigan: 2014.
  37. Parisio MS, Gould JR, Vandenberg JD, Bauer LS, Fierke MK. Evaluation of recovery and monitoring methods for parasitoids released against emerald ash borer. *Biological Control*,2017;106:45-53. doi: <https://doi.org/10.1016/j.biocontrol.2016.12.009>
  38. Halsch CA, Shapiro AM, Fordyce JA, Nice CC, Thorne JH, Waetjen DP, Forister ML. Insects and recent climate change. *PNAS*,2021;118(2):e2002543117. doi: <https://doi.org/10.1073/pnas.2002543117>
  39. Dar AA, Jamal K. Moths as ecological indicators: a review. *Munis Entomology & Zoology*,2021;16(6):8333-839.
  40. Bernardes ACC, Silva RA, Rebelo JMM, Viana JH, Siqueira GM. Abundance and diversity of beetles (Insecta: Coleoptera) in land use and management systems. *Revista Brasileira de Ciencia do Solo*,2020;44:e0190183. doi: <https://doi.org/10.36783/18069657rbc20190183>
  41. Mahanta DK, Singh MK, Sujatha GS, Teja KSS. Insects as Ecological and Biological Indicators. *Vygyan Varta*,2022;3(5):136-143.
  42. Segev N, Groner E, Bouskila, Berger-Tal O. Can microhabitat preferences of ground-dwelling insect be a good indicator for terrestrial ecosystem recover after an oil spill? *Journal Insect Conservation*,2023;23:947-959. doi: <https://doi.org/10.1007/s10841-023-00511-w>
  43. Makwela MM, Slotow R, Munyai TC. Carabid Beetles (Coleoptera) as Indicators of Sustainability in Agroecosystems: A Systematic Review. *Sustainability*,2023;15(5):3936. doi: <https://doi.org/10.3390/su15053936>
  44. Rocha JRMD, Almeida JR, Lins GA, Durval A. Insects as indicators of environmental changing and pollution: a review of appropriate species and their monitoring. *HOLOS Environment*. 1998;10(2):250-262.
  45. Borror DJ, White RE. *A Field Guide to Insects: America North of Mexico*,2nd ed. Houghton Mifflin Company: 1970.
  46. Zaghoul A, Saber M, Gadow S, Awad F. Biological indicators for pollution detection in terrestrial and aquatic ecosystems. *Bulletin of the National Research Centre*,2020;44:127. doi: <https://doi.org/10.1186/s42269-020-00385-x>
  47. Brewer GJ, Boxler DJ, Domingues LN, Fryxell RTT, Holderman C, Loftin KM, Machtinger E, Smythe B, Talley JL, Watson. Horn fly (Diptera: Muscidae) –

- Biology, management, future research directions. *Journal of Integrated Pest Management*,2021:12(1):42. <https://doi.org/10.1093/jipm/pmab019>
48. Boulahia-Kheder S. Advancements in management of major fruit flies (Diptera: Tephritidae) in North Africa and future challenges: A review. *Journal of Applied Entomology*,2021:145(1):939-957. doi: <https://doi.org/10.1111/jen.12938>
49. Roy S, Ghara S, Paria S, Mardaraj PC, Chakraborty SK. Diversity and distribution of true flies (Diptera) of Kuldiha Wildlife Sanctuary, Odisha, India: Functional roles based on ecological guilds. *Entomon*,2023:48(3):397-410. doi: <https://doi.org/10.33307/entomon.v48i3.941>